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Determination of in vitro rumen digestibility and potential feed value of tiger nut varieties

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Abstract

Tiger nut (Cyperus esculentus L.), or chufa, is a plant that is found in nature and is cultivated for its edible tubers. The purpose of this study was to determine the chemical composition, nutritive value, and in vitro digestibility of three tiger nut varieties using the in vitro gas production technique. These varieties were Sarışeker (yellow), Introduction 1, and Balyumru (brown). Rumen fluid was obtained from two cannulated Holstein animals. Time-dependent in vitro gas production was monitored at 3, 6, 9, 12, 24, 48, 72, and 96 hours of incubation. The varieties differed in dry matter (DM), crude ash (CA), ether extract (EE), neutral detergent fibre (NDF), and non-fibre carbohydrate (NFC) content (P < 0.05). They also differed in the instantaneous volume of gas produced and in time-dependent gas production. Balyumru produced more gas at the onset of incubation than Introduction 1 and Sarışeker. However, over time, the gas produced by digestion of Introduction 1 exceeded the other two varieties. The amounts of gas produced at each time-point were intercorrelated. It is recommended that these results should lead to further evaluation in in vivo studies.

Keywords: chemical composition, energy content, in vitro gas production

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Introduction

Farmers that are engaged in stockbreeding in developing countries face problems in the dry season, which include feed shortages, increased feed prices, and climate change. These challenges also affect the quality and quantity of animal protein available for human consumption. To overcome them, it is essential to focus on the use of lesser-known non-traditional feed ingredients such as tiger nuts (*Cyperus esculentus L.*) in animal nutrition (Belewu *et al.*, 2007; Anonymous 2009a; 2009b; Agbabiaka *et al.*, 2013; Suleiman *et al.*, 2018).

Tiger nut (*Cyperus esculentus L.*) or chufa is a plant that is found in nature and is cultivated for its edible tubers, mostly in tropical and temperate regions. Tiger nut tubers may be consumed raw, processed as flour, and used in animal feed production. In addition, they are utilized in the pharmaceutical, cosmetics and perfumery industries (Nazlican, 2007). High-quality biofuel is obtained from tiger nut oil (Bilgili *et al.*, 2018), which is good for human and animal health because it contains 18% saturated fatty acids (palmitic and stearic acids) and 82% unsaturated fatty acids (oleic and linoleic acids) (Zhang *et al.*, 1996). Since tiger nuts do not contain gluten and cholesterol, they are considered a digestive tonic, and thus relieve bloating. Besides, their sodium content is low (Martinez, 2003).

To date, information on the potential use of tiger nuts in ruminant livestock nutrition has been limited. Previous studies focused on its use in feeding poultry, pigs (Ukpabi *et al.*, 2019) and fish. For example, Bamgbose *et al.* (2003) reported that using 33.3% tiger nuts instead of corn in rooster rations reduced feed costs significantly without affecting carcass parameters. Onunkwo and Ugwuene (2015) replaced 0%, 25%, 50%, and 75% corn with tiger nut meal in rations for Japanese quail. After 49 days, they reported that up to 50% of the corn could be replaced with tiger nut meal without reducing the growth of the birds. Similar

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ISSN 0375-1589 (print), ISSN 2221-4062 (online) Publisher: South African Society for Animal Science findings were obtained by Agbabiaka *et al.* (2013) and Obidinma (2009), who reported that tiger nuts could replace 50% of corn in broiler rations. But the use of tiger nuts in feeding ruminant animals was evaluated only by Belewu *et al.* (2007), and that was carried out on goats. Therefore, in this study, the nutrient content, nutritive value, and digestibility in cattle of three tiger nut varieties grown in Turkey were examined with the in vitro gas production technique.

Materials and methods

The experiment was conducted at EMARI (36°51'18" N 35°20'49" E) in Adana Province in Turkey. The tiger nuts were grown at the institute and were collected in three replicates to represent all the samples. The dry feed samples were first ground in a mill with a sieve diameter of 1 mm and then used for the analyses. To determine the dry matter (DM) content, the ground samples were kept in an oven at 70 °C for 24 hours and the differences between the weights before and after baking were computed and expressed in DM%.

Dry matter, CA, CP, and EE were analysed according to the methods described by AOAC (1996). Acid detergent fibe (ADF) and NDF were analysed by the method described by Van Soest *et al.* (1991). Crude cellulose (CC) was determined Pinkerton's (2005) method. Hemicellulose (HC) was calculated with equations by Tekce and Gül (2014). In computing the non-structural carbohydrate (NFC) values, this equation by Weiss *et al.* (1992) was used:

$$NFC \% = 100 - (NDF \% + CP \% + EE \% + CA \%)$$

Total digestibility nutrients, digestible energy (DE), metabolizable energy (ME) and net energy lactation (NEL) were calculated using these formulas (MAFF, 1984):

$$TDN(\%) = 27.66 \ x \ ME \ (Mcal/kg \ DM)$$

$$DE \ (Mcal/kg \ DM) = 0.04409 \ x \ TDN \ (\%)$$

$$ME \ (Mcal/kg \ DM) = [3227 - \frac{\{35.85xADF(\%) + 33.46xCP \ (\%) - 35.85 \ x \ CA(\%)\}}{1000}$$

$$NEL \ (Mcal/kg \ DM) = \{0.0245 \ x \ TDN \ (\%)\} - 0.12$$

To determine the nutrient digestibility of tiger nut varieties through in vitro gas production technique, two Holstein cattle were used, weighing approximately 575 kg, and fitted with rumen cannulas. The experimental animals were housed at Bursa Uludağ University Veterinary Faculty Application and Research Centre. They were fed in two meals, morning and evening, with a mixture of 60% corn silage and 40% concentrate. The animals were provided continuously with clean water.

In vitro gas production from digestion of tiger nuts was determined using the gas production technique (Menke & Steingass,1988) with 100 mL glass syringes (Model Fortuna, Häberle Labortechnik, Lonsee-Ettlenschieβ, Germany). Rumen liquid was collected just before the morning feeding and brought to the laboratory in a vacuum flask. It was filtered through three layers of cheesecloth. Approximately 200 mg of a feed sample was weighed and combined with 40 ml of an incubation medium in three replicate syringes. In addition to the rumen fluid, the incubation medium consisted of 620 ml pure water, 310 ml macro element solution, 0.16 ml trace element solution, 310 ml buffer solution and 1.6 ml resazurin and reduction solutions. After being prepared for digestion the syringes were maintained in a water bath at 39 °C.

The volume of the gas formed in the syringes was recorded after 3, 6, 12, 24, 48, 72, and 96 hours of incubation. The gas production data were modelled with an exponential equation (Ørskov & McDonald, 1979):

$$GP = a + b (1 - e^{-ct})$$

using the NeWay computer program (Chen, 1994) in which:

GP = time-dependent (t) gas production from the substrate (mL)

a = instantaneous volume of the gas produced as the feed is placed in the artificial rumen (mL)

b =time-dependent volume of the gas formed (mL)

a + b = total (potential) gas production (mL)

c = gas production rate constant (hour⁻¹ or %)

t = time (hour)

Analysis of variance procedures (SPSS Inc., Chicago, Illinois, USA) were used to test differences in digestibility and parameter estimates from the gas production curve among the tiger nut varieties after 3, 6, 12, 24, 48, 72, and 96 hours of in vitro digestion. Tukey's multiple comparison test was used to assess differences between varieties. Correlations between digestibility and the parameter estimates for the gas production curves at the 3rd, 6th, 12th, 24th, 72nd, and 96th hour of incubation were calculated as a measure of their relationship.

Results and Discussion

Differences in DM, CA and EE contents between varieties were statistically significant. The CP contents of these varieties were not detected as being different (P > 0.05). In fibre constituents, only the NDF content was found to vary. The more rapidly degradable NFC fraction also differed among varieties, and Sarışeker had with a markedly lower value than Introduction 1 and Balyumru. Values for the nutrient contents of the three varieties of tiger nuts are given in Table 1.

Table 1 Nutritional content of three tiger nut varieties, as a percentage of dry matter

Nutrient, %	Varieties				Dyalia
	Sarışeker	Introduction 1	Balyumru	- SE	<i>P</i> -value
Dry matter	94.62 ^a	93.86 ^b	93.75 ^b	0.172	<0.001
Crude ash	2.65 ^a	2.52 ^b	2.42 ^c	0.044	0.005
Crude protein	4.45 ^a	4.41 ^a	4.32 ^a	0.127	0.942
Ether extract	28.43 ^a	25.84 ^b	20.28 ^c	1.523	<0.001
Acid detergent fibre	14.91 ^a	14.01 ^a	16.21 ^a	0.484	0.168
Neutral detergent fibre	23.86 ^b	21.72 ^c	26.51 ^a	0.882	0.001
Acid detergent lignin	5.64 ^a	5.77 ^a	5.74 ^a	0.140	0.960
Crude cellulose	9.26 ^a	8.24 ^a	10.47 ^a	0.516	0.229
Hemi cellulose	8.94 ^a	7.71 ^a	10.30 ^a	0.537	0.103
Non-structural carbohydrate	40.61 ^b	45.51 ^a	46.47 ^a	1.152	0.001

 $[\]overline{a}$, within a row values with a common superscript were not different at \overline{P} =0.05

The DM content of tiger nut tubers for varieties in Spain ranged from 91.3% to 92.9% and from 93.1% to 93.4% for the varieties in Turkey (Thomas, 2014). Aduwamai *et al.* (2018) reported that the DM contents of unprocessed, soaked, and roasted tiger nuts were 90.63%, 90.63%, and 93.67%, respectively, and the effect of processing techniques on DM was statistically significant. Emurotu (2017) stated that DM content in tiger nuts varied and was 88% in yellow tiger nuts and 89.98% in brown tiger nuts. Bobreneva & Baioumy (2018) found the DM content of tiger nuts was 91.70%, whereas Madaki *et al.* (2018) stated that the DM contents of processed and unprocessed tiger nuts were 92.11% and 90.33%, respectively. These values were lower than the DM values that the authors found in the current study because of the varieties, the climate and soil structure of the region, and the processing techniques.

Tiger nut lines grown in the Çukurova region of Adana, southern Turkey, contained an average of 93.28% DM, 24.5% EE, 25.68% starch, 1.42% CA, 5.05% CP, and 8.91% CC. Özcan *et al.* (2010) found that the CP, EE, CC, and CA levels of tiger nut tubers were 8.11%, 21.60%, 22.13%, and 2.34%, respectively. The CP content found by Özcan *et al.* (2010) was substantially higher than the CP levels that were observed in this study. Gambo and Da'u (2014) stated that the CP content of tiger nuts varied based on strain and drying, and that the CP content of yellow tiger nuts (7.15%) was lower than that of brown tiger nuts (9.70%). Asante *et al.* (2014) stated that the CP content of varieties ranged between 4.62% and 5.46% on average. Bado *et al.* (2015) stated that the CP content of tiger nuts varied between 3.30 and 4.33 g/100 g, a result that is more similar to the present findings. The variance observed in the CP results of tiger nuts in several studies may stem from the strains or lines, the regions in which the crop was grown, analysis methods, processing techniques, and harvest times.

Asante *et al.* (2014) found that the fat content varied significantly among varieties from 19.27% to 21.92%, values that are most similar to the EE content of Balyumru in the present study. Bado *et al.* (2015)

reported that the EE content of tiger nut tubers varied between 24.91 and 28.94 g/100 g, results that are more similar to the present findings for Sarışeker and Introduction 1. Gambo and Da'u (2014) reported that the EE content in tiger nuts varied depending on strain and drying, and ranged between 32.13% and 35.43%. Thomas (2014) asserted that the EE content was between 23% and 28.3%. Achoribo and Ong (2017) stated that it was 30%. As Ezeh *et al.* (2014) reported, the variations in tiger nut oil content resulted from the origin of the tuber, region, genetic structure, variety, and age of the tissue. Emurotu (2017) found the EE content in tiger nuts ranged from 21.9% to 23.1% with an average value of 22.45%. Madaki *et al.* (2018) stated that the EE content of processed and unprocessed tiger nuts varied between 25.53% and 28.61%. Oleic acid, palmitic acid, and linoleic acid are major contributors to the oil extracted from tiger nuts (Coşkuner *et al.*, 2002). Thus, it can be concluded that tiger nuts are an energy dense feedstuff. However, high level of fats in the diet could prove toxic to rumen microbes and affect fibre digestibility (Behan et al., 2019). Thus the fat content of tiger nuts should be considered in formulating rations for ruminant animals when tiger nuts are fed.

The average ADF content, an indicator of digestibility of roughages (Van Soest, 1967), was 15.04% in this study. Balyumru had the highest ADF content with 16.21%. When the NDF contents were examined, the lowest NDF was found in Introduction 1 with 21.72%, whereas the highest NDF content was observed in Balyumru with 26.51%. The neutral detergent fibre content is closely related to the feed consumption. The recommended ADF and NDF contents in the total ration for dairy cattle are 23% and 36 - 38%, respectively. The ideal NDF content from the roughage is accepted to be 27 - 28% (NRC, 2001).

Asante *et al.* (2014) reported that the CC of several varieties ranged between 7.42% and 11.8% and between 7.5% and 13.5%. Thomas (2014) stated that the CC for the tiger nut tubers of the varieties in Spain ranged between 9.8% and 11.0% and that it was 8.7% for the varieties in Turkey. Emurotu (2017) found the CC content in tiger nuts was between 23.3 and 26.0% with an average of 24.7%, while Bobreneva and Baioumy (2018) found it was 15.37%, and Madaki *et al.* (2018) observed values between 22.34% and 24.94%. Based on the results pertaining to NDF, ADF, and CC ruminant animals fed tiger nut tubers will likely require supplementation with other sources of fibre.

Non-fibre carbohydrate (NFC) content is calculated through a formula that uses NDF, CP, EE, and CA (Table 4). A statistically significant difference was found between the varieties in terms of NFC, and the average NFC content was 44.20%. The total digestible nutrients ranged from 71.59% to 73.78%, revealing a statistically insignificant difference (Table 4) (*P* >0.05).

Beyond the initial gas production recorded at three hours of incubation, no further differences were observed until the 24th hour (Table 2). The highest gas production at that point was observed in Introduction 1, whereas Sarışeker and Balyumru had produced significantly less gas at that time. This pattern of Introduction 1 producing more gas than the other two varieties was maintained through 96 hours. For all three varieties, the gas production reached stasis at approximately 24 h which might be interpreted as implying an impairment of microbial digestion by the high fat content of tiger nuts.

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Varieties	3 h	6 h	12 h	24 h	48 h	72 h	96 h
Introduction 1	16.71 ^a	22.43 ^a	33.18 ^a	42.76 ^a	46.05 ^a	48.77 ^a	56.44 ^a
Sarışeker	11.38 ^b	15.43 ^a	24.85 ^a	34.93 ^b	35.68 ^b	36.44 ^b	36.60 ^b
Balyumru	12.04 ^b	16.76 ^a	26.85 ^a	34.10 ^b	35.52 ^b	37.27 ^b	39.10 ^b

1.80

0.134

2.07

0.025

2.34

0.022

3.47

0.007

1.72

0.044

Table 2 In vitro gas production of three tiger nut varieties as a function of incubation time

1.49

0.110

1.07

0.059

SE

P-value

Initial gas production was higher for Balyumru than for Introduction 1 and Sarışeker (Table 3). However, over time the gas produced from Introduction 1 exceeded that produced by the other two varieties. The rate parameter did not differ. Thus, the net effect of less gas being produced by Sarışeker was an indication of a possible lower degree of ruminal fermentation.

Although there were significant differences among the tiger nut varieties, the predicted energy content for ruminant livestock, whether measured as total digestible nutrients (TDN), DE, ME, or NEL, did not differ significantly in in vitro gas production (Table 4). This was contrary to the expectation based on Menke and

 $^{^{}a,b}$ Within a column values with a common superscript were not different at P = 0.05

Steingass (1988) of a strong correlation between ME and 24 hours in vitro gas production. The metabolizable energy content of tiger nuts differed, depending on the variety, and ranged from 4.59 to 4.71 Mcal/kg (Gambo & Da'u, 2014).

Table 3 Parameter estimates for exponential equation describing kinetics of in vitro gas production of three varieties of tiger nuts through 96 hours of incubation

Varieties	а	b	С	a+b
Introduction 1	2.5 ^b	44.87 ^a	0.10 ^a	47.37 ^a
Sarışeker	2.48 ^b	34.92 ^b	0.09 ^a	37.40 ^b
Balyumru	3.70 ^a	39.51 ^b	0.08 ^a	43.21 ^a
SE	0.22	1.60	0.01	1.61
P-value	0.004	0.007	0.235	0.007

¹Ørskov & McDonald (1979)

Table 4 Estimated total digestible nutrients, digestible energy, metabolizable energy and net energy for lactation contents of three tiger nut varieties

Varieties	TDN, % DM	DE, Mcal/Kg DM	ME, Mcal/Kg DM	NEL, Mcal/Kg DM
Sarışeker	72.98 ^a	3.22 ^a	2.64 ^a	1.67 ^a
Introduction 1	73.78 ^a	3.25 ^a	2.67 ^a	1.69 ^a
Balyumru	71.59 ^a	3.16 ^a	2.59 ^a	1.63 ^a
SE	0.468	0.020	0.017	0.011
P-value	0.077	0.123	0.123	0.123

^a Within a column values with a common superscript letter did not differ at P = 0.05

The amounts of gas produced by in vitro incubation of tiger nuts were highly correlated (P > 0.85) across time (Table 5). Thus, a 96 hours of in vitro incubation may have been longer than necessary.

Table 5 Estimates of Pearson correlation among quantities of gas produced over time from in vitro incubation of tiger nuts

Time	6 h	12 h	24 h	48 h	72 h	96 h
3 h	.966**	.937**	.841**	.911**	.922**	.868**
6 h		.991**	.846**	.895**	.901**	.876**
12 h			.860**	.894**	.882 ^{**}	.859**
24 h				.980**	.919 ^{**}	.893**
48 h					.971**	.926**
72 h						.964**

^{**} *P* <0.01

a,b Within a column values with a common superscript were not different at P=0.05

a: instantaneous gas production (mL), b: time-dependent volume of gas produced (mL), c: gas production rate constant

TDN: total digestible nutrients, DE: digestible energy, ME: metabolizable energy, NEL: net energy for lactation, DM: dry matter

In examining the correlations among parameter estimates (Ørskov & McDonald, 1979), the large correlation (0.991) between estimates for the volume of gas formed over time and the total amount of gas produced indicated that the instantaneous gas production was negligible. It seems that the time-dependent volume of the gas explained virtually all of the total gas production. None of the other correlations among the estimated parameters were significant (P > 0.05), although the correlation between the instantaneous volume of gas produced and the gas production rate constant (-0.355) was of a magnitude to merit further study.

Conclusion

Tiger nuts may be regarded as an energy feed owing to their high fat content and abundance of carbohydrates. Thus, tiger nuts may have potential as an inexpensive source of energy for feeding ruminant animals. If tiger nuts are used in livestock feeding, this may reduce the competition between humans and animals for maize and other cereals. The use of tiger nuts in the diets for ruminant animals needs further study. The results from this research should be supported by an in vivo feeding and digestion trials.

Authors' Contributions

TA contributed to the project idea, design and execution of the study. ES, İÜ, HH, PÇ conducted the laboratory analyses. TA, ES and BDÖ supervised the experiment and wrote the manuscript.

Conflict of Interest Declaration

The authors declare that that there are no competing interests.

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